



Extrachromosomal circular DNA-based amplification and transmission of herbicide resistance in crop weed *Amaranthus palmeri*

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Gene amplification has been observed in many bacteria and eukaryotes as a response to various selective pressures, such as antibiotics, cytotoxic drugs, pesticides, herbicides, and other stressful environmental conditions. An increase in gene copy number is often found as extrachromosomal elements that usually contain autonomously replicating extrachromosomal circular DNA molecules (eccDNAs). *Amaranthus palmeri*, a crop weed, can develop herbicide resistance to glyphosate [N-(phosphonomethyl) glycine] by amplification of the 5-enolpyruvylshikimate-3-phosphate synthase (EPSPS) gene, the molecular target of glyphosate. However, biological questions regarding the source of the amplified EPSPS, the nature of the amplified DNA structures, and mechanisms responsible for maintaining this gene amplification in cells and their inheritance remain unknown. Here, we report that amplified EPSPS copies in glyphosate-resistant (GR) *A. palmeri* are present in the form of eccDNAs with various conformations. The eccDNAs are transmitted during cell division in mitosis and meiosis to the soma and germ cells and the progeny by an as yet unknown mechanism of tethering to mitotic and meiotic chromosomes. We propose that eccDNAs are one of the components of McClintock's postulated innate systems [McClintock B (1978) *Stadler Genetics Symposium*] that can rapidly produce soma variation, amplify EPSPS genes in the sporophyte that are transmitted to germ cells, and modulate rapid glyphosate resistance through genome plasticity and adaptive evolution.

5-enolpyruvylshikimate-3-phosphate synthase | eccDNA | glyphosate resistance | adaptive evolution | gene amplification

Eukaryotic cells use gene amplification mechanisms to over-express specific genes for survival under stress. Amplified gene copies are often found as part of autonomously replicating extrachromosomal circular DNA molecules (eccDNAs), including double minutes (DMs) (1–8). The eccDNAs have been widely observed in many drug-resistant and tumor cell lines. The eccDNAs vary in size, ranging from a few hundred kilobases to megabases (9, 10). The eccDNAs may be simple (oligomeric) in structure, derived without any rearrangement from the corresponding chromosome (11, 12), or complex eccDNAs with duplicated copies of the same gene (13–15). EccDNAs containing sequences from different chromosomal loci have also been reported (16), indicating that different mechanisms may drive eccDNA assembly and evolution.

Despite the lack of centromeres, eccDNAs can be transmitted to daughter cells by tethering of their chromatin body to the telomeric region of segregating chromosomes from anaphase to telophase (17, 18). All reported cases of eccDNAs have been studied in cell lines; their genesis, behavior, and inheritance have not been studied in soma and germ cells of living organisms.

Living organisms, including plants and insects, have also evolved resistance to xenobiotics compounds, such as herbicides and insecticides, via gene amplification (19–25). In all reported studies, amplified gene copies were located in specific chromosomes

or multiple chromosomal regions, but not in the form of eccDNAs (19, 23, 24, 26). Therefore, gene amplification in these living organisms is thought to have occurred by chromosomal rearrangements, such as inversion and reciprocal exchanges, or in association with transposable elements (23, 27–31).

A 40-fold to >100-fold amplification of the 5-enolpyruvylshikimate-3-phosphate synthase (EPSPS) gene is associated with glyphosate resistance in *Amaranthus palmeri* populations (23). Initial reports suggested that the EPSPS amplicon was at least 30 kb in length and contained miniature inverted-repeat transposable elements (MITEs), which were postulated to disperse the amplicon to all of the glyphosate-resistant (GR) *A. palmeri* chromosomes at multiple sites (23, 31). More recently, the length of the EPSPS amplicon was extended to 297 kb, and termed the “EPSPS cassette,” by sequencing overlapping large-insert clones derived from a bacterial artificial chromosome (BAC) library (29). These clones flank the EPSPS gene, which was unique to GR *A. palmeri* across the United States, suggesting a single origin (29, 32). Here, we report that the EPSPS cassette is, in fact, an extrachromosomal circular DNA carrying the EPSPS gene, hereafter referred to as eccDNA. We report on the dynamics of eccDNA structure, variation, and behavior in mitotic and germ cells, as well as possible modes of inheritance, and discuss how they may trigger the plasticity of the GR response.

Significance

Glyphosate is a nonselective herbicide used around the globe for weed control in glyphosate-resistant (GR) and noncrop situations. The extensive and exclusive use of glyphosate has led to the evolution of herbicide resistance in many crop weeds. The molecular target of glyphosate, the 5-enolpyruvylshikimate-3-phosphate synthase (EPSPS) gene, confers resistance upon amplification and was first documented in GR *Amaranthus palmeri*. We now report that amplified EPSPS copies in GR *A. palmeri* are present in the form of extrachromosomal circular DNA molecules (eccDNAs) with various conformations. We discovered that eccDNAs are transmitted to the next generation by tethering to mitotic and meiotic chromosomes. These results represent a report of extrachromosomal structures that drive rapid adaptive evolution in higher organisms.

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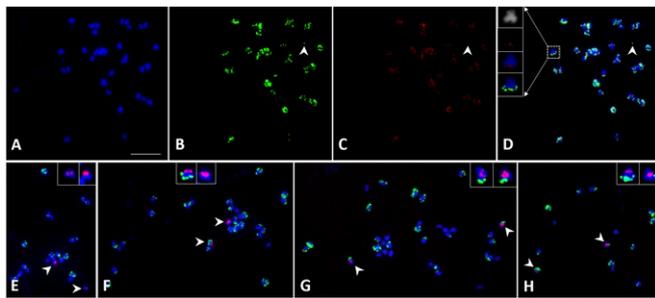


Fig. 1. FISH mapping of eccDNAs and the *EPSPS* gene in mitotic metaphase chromosomes of root meristem cells of GR *A. palmeri* with 80 *EPSPS* copies. (A) Mitotic metaphase chromosomes stained by DAPI. (B) EccDNAs showing green signals overlying all chromosomes except one (arrowhead). (C) All eccDNAs seen in B carry *EPSPS* copies (red signal). (D) Merged image. (Insets) Colocalization of eccDNA (green) and *EPSPS* (red) signals on the tip of a chromosome, but not as part of the chromosome. Arrowheads in B–D point to eccDNA that is not associated with metaphase chromosomes. (E) Marker chromosome labeled with 5S rDNA (red signals) free of eccDNAs. (F) Another cell showed one marker chromosome free of signals, but its homolog has eccDNA. (G) Both marker chromosomes have eccDNA signals. (H) CNV of eccDNA signals in the two marker chromosomes. (E–H, Insets) 5S rDNA-bearing chromosomes showing random distribution of eccDNAs in different cells. Arrowheads in E–H point to 5S rDNA bearing chromosomes showing random distribution of eccDNAs in different cells. (Scale bar, 10 μ m).

Results

Copy Number Variation in the *EPSPS* Gene Is Associated with Unique Chromosome Organization of the *EPSPS* Cassette. We identified glyphosate-sensitive/susceptible (GS) and GR isolates of *A. palmeri* with various *EPSPS* copy numbers ranging from 1 to 120 based on quantitative PCR (qPCR) assays (Table S1). To study chromosome organization in relation to copy number variation (CNV) of the *EPSPS* gene, a DNA probe specific to the *EPSPS* gene and an *EPSPS*-containing BAC 22F22, or its flanking BACs, 5K07 and 1A02, were co-hybridized to chromosomes from several GS and GR *A. palmeri* plants (Fig. 1 and Fig. S1). FISH using an *EPSPS* gene probe on mitotic cells of GS *A. palmeri* with one copy of *EPSPS* revealed a tiny hybridization signal in the pericentromeric region of one pair of chromosomes (Fig. S1 C and D). FISH using BAC 22F22, in addition to a tiny signal on one pair of chromosomes, also generated dispersed FISH sites on all chromosomes from the same GS plant (Fig. S1 B and D).

In an *A. palmeri* plant with 12 *EPSPS* copies, the *EPSPS*-FISH signals on one pair of chromosomes were brighter than those from GS plants with a single copy of *EPSPS*, indicating that the *EPSPS* gene in this plant was amplified near its original location (Fig. S1 G and H). The BAC 22F22-FISH hybridization signals were faint but dispersed over all of the chromosomes except for the significantly more intense hybridization signals at the amplified *EPSPS* gene locus (Fig. S1 F and H).

In a GR *A. palmeri* plant with 80 *EPSPS* copies, *EPSPS*-FISH signals were detected on most chromosomes (Fig. 1). However, the FISH signals appeared to be at the edges of or outside the condensed chromosomes (Fig. 1 C and D). Strong and distinct hybridization signals were generated using BAC 22F22 (Fig. 1B) and were colocalized with those of *EPSPS* signals (Fig. 1D). Hybridization signals not associated with chromosomes were often observed in different metaphase cells of GR *A. palmeri* plants (arrows in Fig. 1 B–D). These data indicate that these structures were probably part of the *EPSPS* cassette reported by Molin et al. (29), and we began studies on the structure of the cassette using FISH on extended DNA fibers.

***EPSPS* Cassette Is an eccDNA Displaying Unique Structural Polymorphisms.** Molin et al. (29) reported the *EPSPS* cassette, which is 297 kb in length, consisting of seven overlapping BACs. Further selection by sequencing of two additional BACs revealed overlapping sequencing of the free ends indicating a potentially circular orientation of

the *EPSPS* cassette. Six BACs associated with the *EPSPS* cassette were used in fiber-FISH mapping. These BACs were grouped into two pools, and they were labeled with alternate green/red colors based on their location in the *EPSPS* cassette (Fig. S2). Surprisingly, we found that ~50.2% of fibers are in circular form, indicating that the *EPSPS* cassette is, in fact, an eccDNA (Fig. 2A, Table 1, and Fig. S2C). Based on the proportion of red and green signal tracks in circular molecules, we consider these eccDNAs to be intact and the wild-type form (Fig. 2A). The microscopic size of the circular form of these eccDNAs varied from ~30 μ m to ~200 μ m, which might be due to the variation of DNA fiber extension in the experiments. Therefore, we were unable to use the microscopic measurement data in classifying the eccDNA types. Instead, we scored eccDNAs for structural polymorphisms based on circular or linear structure and the number and proportion of red and green signals. In the circular DNA class, another 11.8% were determined to be a dimerized circular form of wild-type eccDNA with head-to-tail tandem duplication (hereafter, dimeric eccDNA) (Fig. 2C).

The remaining 38% of the eccDNA showed a linear structure (Table 1). Linearized fibers with different breakpoints but similar in composition to wild-type eccDNA were the most frequent (21.8%) class (Fig. 2B and Fig. S2D). Linear forms of dimeric eccDNAs were also detected (8.0%) (Fig. 2D). We also detected atypical fibers where the hybridization patterns deviated from the expected FISH patterns in 8.2% of the total fibers analyzed (Fig. 2E). Overall, our results demonstrated that ~50% of eccDNAs of GR *A. palmeri* were structurally diverged due to duplication and deletion events (Table 1).

EccDNAs Display CNV and Random Chromosome Associations in Soma Cells of GR *A. palmeri*. The GR *A. palmeri* plants with 80 *EPSPS* copies as determined by qPCR displayed surprising CNV in soma cells as revealed by FISH. The hybridization patterns of the eccDNAs on metaphase cells varied from cell to cell in the same plant. To determine whether the hybridization patterns of metaphase chromosomes were random, a 5S rDNA probe was used in FISH, which showed hybridization signals on one chromosome pair of *A. palmeri*. We observed four different patterns of the eccDNA signals on 5S rDNA-labeled homologous chromosome pairs in different cells ($n = 24$) from a single root tip meristem: (i) Both chromosomes were without eccDNA signals (16.7%) (Fig. 1E), (ii) one of the two chromosomes was without eccDNA signal (25%) (Fig. 1F), (iii) both chromosomes had similar signal intensity (33.3%) (Fig. 1G), and (iv) the two chromosomes varied in signal intensity (25%) (Fig. 1H). We conclude from these data that most of the eccDNAs are extrachromosomal elements that are randomly anchored to the chromosomes at mitotic metaphases.

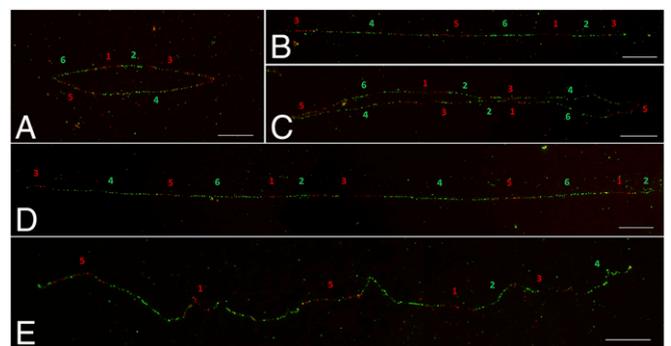


Fig. 2. Fiber-FISH images of eccDNAs in GR *A. palmeri* with 80 *EPSPS* copies. (A) Circular form of eccDNA. (B) Linear form of eccDNA. (C) Dimerized circular form of eccDNA with head-to-tail tandem orientation. (D) Linear form of eccDNA with head-to-tail dimer. (E) Atypical fiber representing structural changes. Note: In the relatively long DNA fibers (D, E), two images were captured sequentially with an overlapping region and then they were combined into a single image using Adobe Photoshop. 1, BAC 01G15; 2, BAC 13C09; 3, BAC 22F22; 4, BAC 23A10; 5, BAC 03A06; 6, BAC 08H14. (Scale bars, 10 μ m).

Table 1. Frequency of different structure polymorphisms of eccDNAs detected by fiber-FISH

Structure	Circular		Dimeric		Atypical*
	Circular	Linear	circular	linear	
No. of observations	564	245	133	90	92
Frequency, %	50.2	21.8	11.8	8.0	8.2

*Fiber-FISH patterns that cannot be discriminated from other four types.

EccDNAs Display Unique Behavior and a Chromosome Tethering Mechanism for Inclusion in Daughter Cells During Meiosis. We analyzed the distribution of eccDNAs in meiotic pachytene chromosomes of GS and GR *A. palmeri* plants (Fig. 3). As expected, the eccDNAs were not observed in GS *A. palmeri* plants with one to 12 *EPSPS* copies (Fig. 3 E–H). However, 4',6'-diamidino-2-phenylindole (DAPI)-stained pachytene chromosome of the GR *A. palmeri* revealed numerous eccDNAs outside the chromosome axis (arrowheads in Fig. 3A), indicating that eccDNAs were not integrated into the chromosomes. The BAC 22F22 and *EPSPS* gene signals were colocalized (Fig. 3 B–D).

To study this apparent tethering of the eccDNAs to chromosomes during cell division further, we analyzed eccDNA behavior during all stages of meiosis I and II from leptotene to telophase II and also in immature pollen grains (Fig. 4 A–J). Numerous eccDNAs can be seen associated with leptotene and zygotene chromosomes, and a few are lying in the cytoplasm (arrowheads in Fig. 4 A and B). At pachytene stage, homologous chromosomes are fully paired. If eccDNAs were integrated into the chromosomes, then double signals would be observed; however, most signals were not double but, instead, single or in clumps and lying next to the chromosomes (Fig. 4C). Moreover, random and variable association of eccDNAs with different chromosomes was seen in well-spread chromosomes at pachytene (Fig. 4C), diplotene (Fig. 4D), diakinesis (Fig. 4E), and metaphase stages (Fig. 4F). The association of eccDNAs with laggard and stretched chromosomes was clearly observed at anaphase I (Fig. 4G) and anaphase II (Fig. 4I). By metaphase II, a few eccDNAs were seen lying away from the chromosomes in the cytoplasm (Fig. 4H). Pollen from GR plants also showed eccDNA signals, indicating transmission to the gametophyte (Fig. 4J).

EccDNAs Are Sexually Transmitted to Progeny Plants and Display Dramatic CNV in Soma Cells. Our meiotic chromosome study indicated the possibility of transmission of the eccDNAs to offspring by a chromosome tethering mechanism. To study sexual transmission, we made crosses between a female GS *A. palmeri* plant lacking eccDNAs and a male GR *A. palmeri* plant carrying the eccDNAs, and vice versa. Plants grown from the seed of these crosses were labeled F₁ plants (F₁). Ten F₁ plants from each reciprocal cross were randomly selected for qPCR and FISH analysis (Fig. 5 and Fig. S3). FISH analyses of root tip cells in these 20 F₁ plants showed that all of the plants had positive signals associated with their mitotic metaphase chromosomes, indicating transmission of eccDNAs to the offspring (Fig. 5 and Fig. S3).

We found that the eccDNA in F₁ progeny displayed CNV ranging in number from one to 39 (Fig. 5F and Fig. S3A). All progeny except one harbored eccDNAs, but FISH signals on mitotic metaphase spreads prepared from a single root tip meristem were dramatically variable from cell to cell (Fig. 5). We observed several FISH signal patterns in different cells ($n = 50$) from a single root preparation of one plant [male highly glyphosate resistant and female glyphosate susceptible (MHFS 1)]: (i) eccDNAs were associated with most of the chromosomes in 50% of the cells similar to the GR parent plant (Fig. 5 A and B), (ii) eccDNAs were associated with half of the chromosomes in 26% of the cells (Fig. 5C), (iii) eccDNAs were associated with only a few of the chromosomes in 16% of the cells (Fig. 5 A and D), and (iv) all chromosomes were free of eccDNAs in 8% of the cells (Fig. 5E).

Similar FISH patterns were detected on the metaphase cell spreads in all 20 F₁ plants (Fig. 5 A–E and Fig. S3). These results showed that eccDNAs varied in copy number due to missegregation during mitotic divisions and were not integrated into chromosomes.

EccDNAs Display CNV in Different Tissues of a Plant. Most surprisingly, qPCR analysis using genomic DNA prepared from leaf tissue showed that *EPSPS* copy number in five (MHFS 1, MHFS 2, MHFS 3, MHFS 6, and MHFS 9) of 10 F₁ plants was similar to the copy number found in GS plants (Fig. 5F). However, FISH analysis from root tip meristems indicated that >90% of mitotic metaphase cells in these five plants had positive FISH signals (Fig. 5 A–D).

To resolve this apparent contradiction, we then performed FISH on nuclei isolated from leaf tissue of selected plants used in qPCR analysis (Fig. 5 G and H). The plant MHFS 1 with an estimated one *EPSPS* copy showed positive FISH signals in 12% ($n = 100$) of the nuclei isolated from the leaf tissue cells (Fig. 5G). The plant MHFS 8 with an estimated 31 *EPSPS* copies showed positive FISH signals in 71.4% ($n = 70$) of the nuclei isolated from the leaf tissue cells (Fig. 5H).

Next, we analyzed eccDNA variation in germ cells of plant MHFS 1 with an estimated one copy of the *EPSPS* gene. The results revealed that 12 of 20 cells at the pachytene stage of prophase I of meiosis showed eccDNAs ranging in number from one to 15 and eight of 20 of the cells were lacking eccDNAs (Fig. 5I and Fig. S4). In tapetum tissue, seven of 10 of the cells of this plant had eccDNAs. The frequency of eccDNAs in plant MHFS 1 was higher in the mitotic root tip (92%) and meiotic cells (60–70%) than in cells from leaf tissue (12%); the reasons for this variation are not known except that the former are actively dividing cells, while the leaf is a nondividing, differentiated tissue.

Discussion

This is a report on the role of eccDNA-driven gene amplification and rapid adaptive evolution in higher organisms. The lifestyle of higher organisms, including flowering plants and mammals, alternates between the dominant sporophytic phase and short-lived gametophytic phase. The male and female gametophytes produce the gametes and transmit the genetic information, and the zygotes develop into the sporophytes. The Darwinian evolution model acts on random, preexisting genetic variation in individuals and populations. In Darwinian evolutionary theory, there is no role of the life experience of the sporophyte or the inheritance of acquired characters as Lamarck (33) had proposed. McClintock (34, 35), based on her research on chromosome structure and behavior in soma and germ cells of maize, proposed that sporophytic genomes, in fact, can respond to challenges, such as stress, and that this acquired genomic variation is transmitted to the germ cells. McClintock

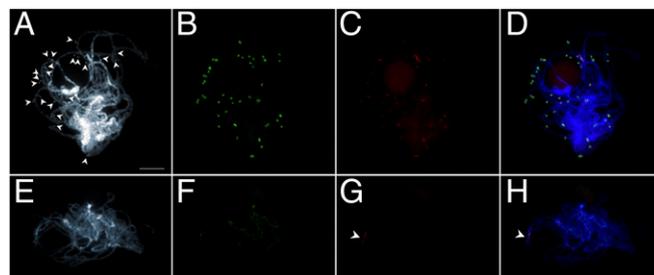


Fig. 3. FISH mapping of eccDNA (green signals) and the *EPSPS* gene (red signals) on meiotic pachytene chromosomes of GR *A. palmeri* with 80 *EPSPS* copies (A–D) and GS *A. palmeri* with 12 *EPSPS* copies (E–H). (A) DAPI-stained pachytene chromosome showing eccDNAs lying outside the pachytene chromosomes (arrowheads). (B) FISH signals with the eccDNA probe. (C) FISH signals with the *EPSPS* gene probe. (D) Colocalization of eccDNA and *EPSPS* probes to eccDNAs. (E) DAPI-stained pachytene chromosome of GS *A. palmeri* did not reveal the eccDNAs. There was faint dispersed signals with eccDNA probe (F), but there was an amplified *EPSPS* gene signal in the pericentromeric region of one chromosome pair (G and H, arrowheads). (Scale bar, 10 μ m.)

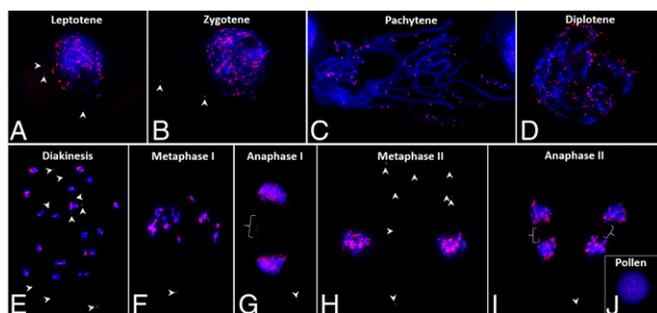


Fig. 4. Distribution of eccDNAs (red signals) on meiotic chromosomes in microsporocytes of GR *A. palmeri* during progression from the leptotene stage of meiosis I through anaphase of meiosis II (A–I) and pollen (J) detected by FISH (arrowheads point to the eccDNAs that are not associated with chromosomes). Brackets in G and I represent the lagging eccDNAs associated with chromatin bridges at anaphase to telophase stages.

(34) proposed the “...presence of innate systems that are able to restructure a genome... to be triggered into action by form of stress... according to the nature of the challenge...”. We propose that eccDNA elements identified in this research are one component of McClintock’s postulated innate system that rapidly produced soma variation, drove amplification of *EPSPS* genes in the sporophyte, and were transmitted to germ cells and modulated rapid evolution of glyphosate resistance in *A. palmeri*.

Wahl (13) proposed a general role of eccDNAs in gene amplification in mammalian and rodent cell lines for many different genes and selective drug agents. He proposed that eccDNAs may originate from chromosomes by deletions or circularization of blocked replicative forks, grow into DMs that are visible under the microscope, undergo unequal segregation during mitotic divisions in the presence of selective agents, and may integrate into the chromosomes to form homogeneously staining regions. Recent work on human cancer lines using combined whole-genome sequencing and cytogenetic analysis has validated the essential role of eccDNA in oncogene amplification, heterogeneity, and the evolution of cancer (1). In yeast, 23% of the genome is represented in eccDNAs ranging in size from 1 to 38 kb, and 80% of eccDNA contained autonomously replicating sequences (36).

EccDNAs have been documented in many plant species, ranging in size from 2 to 20 kb and containing tandem repeats, suggesting their origin via intrachromosomal homologous recombination (37). Our results support widespread occurrence of eccDNA and its crucial role in gene amplification and plasticity of the sporophytic genome response to challenge.

Initial reports suggested that the *EPSPS* amplicon was at least 30 kb in length and contained MITEs, which were postulated to disperse the amplicon to each of the GR *A. palmeri* chromosomes at multiple sites (23, 31). *EPSPS* amplicon FISH signals were interpreted as being dispersed and integrated throughout the chromosome complement of *A. palmeri* in these studies (23, 29, 31). However, somatic metaphase chromosome-based analysis by Gaines et al. (23) did not provide the resolution to detect the tethering of FISH signals to chromosomes. Using a marker chromosome tagged with 5S rDNA-FISH signal, we observed random association of eccDNA signals to the marker chromosome. If the eccDNA was integrated into the chromosome, then they would display uniform signals in all cells, which was not the case (Fig. 1). Moreover, during the pachytene stages of meiosis when chromosomes are highly elongated, the FISH signals were clearly observed as being associated with, rather than part of, the chromosome, and some were not associated with any chromosome at all (Fig. 3). Finally, unequal mitotic segregation of eccDNAs produced variable FISH signals in different cells in the same preparation, and some were lacking the signal (Fig. 5 and Fig. S3). These data indicated that eccDNAs are not integrated into the chromosome, and are autonomously replicating structures that display unequal mitotic segregation, and thereby produce soma cell heterogeneity for resistance evolution.

Molin et al. (29) prepared a BAC library from a GR *A. palmeri* biotype from Mississippi, and sequencing of overlapping BACs revealed a 297-kb sequence unique to GR *A. palmeri*, which they termed an *EPSPS* cassette. The *EPSPS* cassette consisted of an array of repetitive sequences, 72 putative genes, and an autonomous replication sequence (ARS). *EPSPS* cassette-specific marker analysis revealed that GR biotypes across the United States had a single origin (29, 32). Using overlapping BACs from the Mississippi biotype, our fiber-FISH analysis of a Kansas GR biotype clearly established that the *EPSPS* cassette is, in fact, an eccDNA (Fig. 2 and Fig. S2). The shared common structure also supported a single origin of eccDNA of GR biotypes (32). The discovery of ARSs in eccDNA supports our analysis of eccDNA

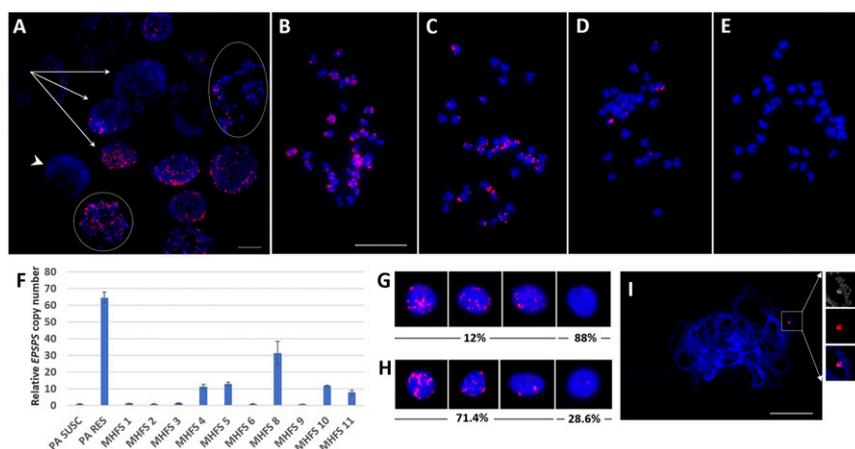


Fig. 5. Inheritance and soma cell heterogeneity of eccDNAs and *EPSPS* copy number in F_1 plants of GS *A. palmeri* \times GR *A. palmeri*, including cells lacking eccDNAs (arrowheads) in a single root tip meristem. (A) Variable number of eccDNA FISH signals (red) in interphase and prometaphase stages (circles). (B–E) Different metaphase cells showing variable numbers of eccDNAs, including no eccDNAs (E). (F) Average *EPSPS* copy numbers of F_1 plants and the controls. The y axis represents the relative β -tubulin/*EPSPS* gene copy number. (G) FISH analysis on nuclei isolated from leaf tissue of plant MHFS 1. (H) FISH analysis on nuclei isolated from leaf tissue of plant MHFS 8 showing variable eccDNA signals. Note that 12% ($n = 100$) (G) and 71.4% ($n = 70$) (H) indicate the percentages of different cells having eccDNA associated with FISH-positive nuclei in plants MHFS 1 and MHFS 8, respectively. (I) Pachytene FISH showing anchoring of an eccDNA on pachytene chromosomes in plant MHFS 1. (Scale bars, 10 μ m.) Inset, Top, 2 \times enlarged and pseudo-colored DAPI image showing association of an eccDNA to pachytene chromosome; Middle, FISH labeled eccDNA; Bottom, merged image of top and middle.

the clone was labeled with digoxigenin-11-deoxyuridine triphosphate (Roche Diagnostics) using a standard nick translation reaction. The clone, maize 5S rDNA (54), was labeled with biotin-16-dUTP (Roche). The BAC clones were labeled with either biotin-16-dUTP or digoxigenin-11-dUTP using a nick translation reaction. Biotin- and digoxigenin-labeled probes were detected with Alexa Fluor 488 streptavidin antibody (Invitrogen) and rhodamine-conjugated antidigoxigenin antibody (Roche), respectively.

Image Analysis. Chromosomes were counterstained with DAPI in Vectashield Antifade solution (Vector Laboratories). The images were captured with a Zeiss Axioplan 2 microscope (Carl Zeiss Microscopy LLC) using a cooled CCD camera CoolSNAP HQ2 (Photometrics) and AxioVision 4.8 software. The final contrast of the images was processed using Adobe Photoshop CS5 software.

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